

**Introduction**

This protocol shows optimum transfection condition using HilyMax in MCF7 cells. To tranfect MCF7 cells in 24-well plate, follow "Optimum Condition for Transfection" and "Transfection Procedure". When using the other vessel, refer to Table 2 and adjust the amounts of cells, medium, DNA and HilyMax in proportion to the relative surface area.

**Important Note**

Optimum Transfection condition is possibly chaged by passage number and culture condition. If transfection efficiency is low by followed this protocol, refer to "Transfected Result by HilyMax" and "Troubleshooting".

**Optimum Condition for Transfection (for 24-well plate)**

Table 1 Optimum condition for tranfection to MCF7 cells

Cell Density	50%	
DNA-HilyMax complex formation	Serum-free medium	30 $\mu$ l
	DNA	1.5 $\mu$ g
	HilyMax	4.5-7.5 $\mu$ l
	Incubation time	15 min
Medium change after transfection	Necessary	

**Transfection Procedure (for 24-well plate)**

**Cell preparation**

- ↓ Adjust the concentration of cells to be 50% confluent in 0.5 ml of growth medium prior to transfection.
- ↓ Inoculate the cell suspension onto the 24-well plate.
- ↓ Incubate cells in CO<sub>2</sub> incubator for 24 hr.

**Transfection**

- ↓ Form the DNA-HilyMax complex
  - Add the serum-free medium(without antibiotics) 30  $\mu$ l/well in a sterile plastic tube
  - Add plasmid DNA 1.5  $\mu$ g/well and mix by gentle pipetting
  - Add HilyMax 4.5-7.5  $\mu$ l/well and mix by gentle pipetting
  - Incubate the mixture of DNA and HilyMax solution at room temperature for 15 minutes
- ↓ Add DNA-HilyMax complex to cells in each well and mix by gentle shaking the plate
- ↓ Incubate cells in CO<sub>2</sub> incubator for 18-48 hr
- ↓ (!) Change the growth medium 4 hours after transfection.

**Assay**

- ↓ Measure protein expression

**Transfection in Various Vessels**

Table 2 Transfection condition in various vessels

Culture Vessel	Culture of Cells		Formation of DNA-HilyMax complex		
	Surface Area	Plating Medium	Serum-free Medium	DNA	HilyMax
96 -well	0.3 cm <sup>2</sup>	0.1 ml	10 $\mu$ l	0.3 $\mu$ g	0.9-1.5 $\mu$ l
24 -well	1.9 cm <sup>2</sup>	0.5 ml	30 $\mu$ l	1.5 $\mu$ g	4.5-7.5 $\mu$ l
12 -well	3.8 cm <sup>2</sup>	1.0 ml	60 $\mu$ l	3.0 $\mu$ g	9.0-15.0 $\mu$ l
6 -well	9.2 cm <sup>2</sup>	2.0 ml	120 $\mu$ l	6.0 $\mu$ g	18.0-30.0 $\mu$ l
35 -mm	8.0 cm <sup>2</sup>	2.0 ml	120 $\mu$ l	6.0 $\mu$ g	18.0-30.0 $\mu$ l
60 -mm	21.0 cm <sup>2</sup>	5.0 ml	300 $\mu$ l	15.0 $\mu$ g	45.0-75.0 $\mu$ l
100 -mm	58.0 cm <sup>2</sup>	15.0 ml	900 $\mu$ l	45.0 $\mu$ g	135.0-225.0 $\mu$ l

**Transfected result by HilyMax**

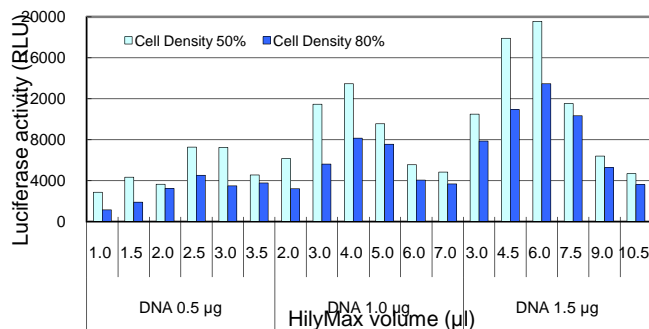


Fig. 1 Transfection Efficiency in MCF7 cells

MCF7 cells were incubated for 24 hr and transfected pGL3 control vector (Promega) using HilyMax in each conditions. Transfection efficiency (Luciferase activity) was measured in 24 hr after transfection. MCF7 cells were cultured in MEM medium(Gibco) containing 10%FBS(Gibco) and Non-Essential Amino Acids (Gibco) for about 2 weeks after thawing.

50% confluent: 0.6 x 10<sup>5</sup> cells/well    80% confluent: 1.0 x 10<sup>5</sup> cells/well

**Troubleshooting**

**-Low Transfection Efficiency-**  
Change the DNA( $\mu$ g):HilyMax( $\mu$ l) ratio to 1:6-1:8. Increase the mass of DNA up to 1.5-2.0 times and change the DNA( $\mu$ g):HilyMax( $\mu$ l) ratio to 1:3-1:5.

**-High cellular Toxicity-**  
Decrease the mass of DNA down to 2/3 and change the DNA ( $\mu$ g):HilyMax( $\mu$ l) ratio to 1:2-1:7.

**-Check the Material and Condition-**  
Was HilyMax Reagent dissolved completely when HilyMax was Prepared?  
Was incubation time of cells after tranfection optimum for cells and plasmid?  
Was DNA-HilyMax complex formed in medium without serum and antibiotics?